EUROPEAN HEALTH AND DIGITAL EXECUTIVE AGENCY (HaDEA)

Department A Health and Food
Unit A2 EU4Health/SMP

Food Programmes for eradication, control and surveillance of animal diseases and zoonoses

submitted for obtaining EU financial contribution

Annex II: Control programme – Reduction of prevalence of Salmonella serotypes in certain poultry populations

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Member state : ESPANA

Disease Salmonella

Animal population Breeding flocks of Gallus gallus

This program is multi annual : no

Request of Union co-financing from beginning : 2023

1. Contact data

Name

Phone

Email

Your job type within the CA :

Submission Date

30/11/2022 13:59:06

Submission Number

1669813152036-18915

Breeding flocks of Gallus gallus

A. Technical information

By submitting this programme, the Member State (MS) attests that the relevant provisions of the EU legislation will be implemented during its entire period of approval, in particular:

- Regulation (EC) No 2160/2003 on the control of *Salmonella* and other specified food-borne zoonotic agents,
- Regulation (EU) No 200/2010 implementing Regulation (EC) No 2160/2003 of the European Parliament and of the Council as regards a Union target for the reduction of the prevalence of *Salmonella* serotypes in adult breeding flocks of *Gallus gallus*,
- Regulation (EC) No 1177/2006 implementing Regulation (EC) No 2160/2003 as regards requirements for the use of specific control methods in the framework of the national programmes for the control of *Salmonella* in poultry.

As a consequence, the following measures will be implemented during the whole period of the programme:

1. The **aim of the programme** is to implement all relevant measures in order to reduce to 1% or less the maximum percentage of adult breeding flocks of *Gallus gallus* remaining positive for the target *Salmonella* serovars: *Salmonella* Enteritidis (SE), Typhimurium (ST)(including the antigenic formula 1,4,[5],12:i:-), Hadar (SH), Infantis (SI) and Virchow (SV).

   For a MS with less than 100 adult breeding flocks of *Galus gallus* the target is to have no more than one such flock remaining positive for the relevant *Salmonella* serovars per year.

☒*yes* ☐*no*

If no please explain.

The objective of the National Programme is to control the presence of the five most frequent serotypes of human salmonellosis: S. Enteritidis, S.Typhimurium, including monophasic strains of Salmonella Typhimurium with the antigenic formula 1,4,[5],12:i:-, S. Virchow and S. Hadar in breeding flocks of Gallus gallus, and to reduce its prevalence to that targeted by the Community, i.e. to a maximum of 1% in flocks with more than 250 adult birds.
Definition of a positive case:
A breeding flock shall be considered positive for the purpose of ascertaining the achievement of the Union target:
a) when the presence of the relevant Salmonella serotypes, other than vaccine strains, has been detected in one or more samples taken from the flock, or
b) when residues of antimicrobials or bacterial growth inhibitors have been detected in the flock.
• A positive breeding flock shall only be counted once regardless of how often the relevant Salmonella serotypes have been detected in this flock during the production period or whether the sampling was carried out at the initiative of the food business operator or by the competent authority. However, if

sampling during the production period is spread over two calendar years, the result of each year shall be reported separately. In the event that a positive result is detected and the competent authority decided to perform a confirmatory analysis, the final valid result shall be the result of the said confirmatory analysis.

2. The programme will be implemented on the **whole territory** of the Member State.

☒*yes* ☐*no*

If no please explain.

3. Flocks subject to the programme

	Total number of flocks of breeders in the MS	Number of flocks with at least 250 adult breeders	Number of flocks where FBO sampling shall take place	Number of flocks where official sampling will take place
Rearing flocks	1 020		1 020	5
Adult flocks	1 720	1 700	1 720	1 700
Number of adult flocks where FBO sampling is done at the hatchery		0	0	0
Number of adult flocks where FBO sampling is done at the holding		1 700	1 720	1 700

NB : All cells shall be filled in with the best estimation available.

Comments (max. 32000 chars) :

It will be implemented in all holdings of Gallus gallus breeding hens (both adult breeding and rearing hens). On breeding hen holdings where the producer directly supplies small quantities of primary products to the final consumer or to local retail establishments directly supplying the primary products to the final consumer, at least one FBO control should be done per year in all the flocks present in the farm at that moment (for harmonisation purposes). The competent authorities of the Autonomous Communities shall take any action required to ensure control and monitoring of salmonellosis with public health significance. This programme will not be implemented at holdings that produce primary products for own consumption (for private domestic use). Holdings to which the programme will apply must be authorised and registered by the competent authorities. For the purposes of the programme an epidemiological unit shall be considered to be a breeding flock, defined as all poultry of the same health status kept on the same premises or within the same enclosure; in the case of housed poultry, this includes all birds sharing the same airspace, in accordance with Article 2(3)(b) of Regulation (EC) No 2160/2003 of the European Parliament and of the Council. Flocks of breeding hens shall be identified individually. To identify the flocks on a holding the REGA code will be used, consisting of a capital letter

corresponding to the shed (this letter must be written on the entrance door to the shed) and the date of entry of the birds into that shed, in the format mmyyyy. REGA+ SHED (CAPITAL LETTER)+ DATE OF ENTRY OF BIRDS (mmyyyy)

4. Notification of the detection of target *Salmonella* serovars

A procedure is in place which guarantees that the detection of the presence of the relevant *Salmonella* serotypes during sampling at the initiative of the food business operator (FBO) is notified without delay to the competent authority by the laboratory performing the analyses. Timely notification of the detection of the presence of any of the relevant *Salmonella* serotypes remains the responsibility of the food business operator and the laboratory performing the analyses.

☒ *yes* ☐ *no*

If no please explain.

Any natural or legal person, especially veterinarians, must notify the competent authorities of any confirmed (or suspected) cases of salmonellosis, regardless of whether or not they are related to measures in the framework of the national programmes for the control of salmonella. To that end, all confirmed or suspected results from samples taken and analysed by operators outside the framework of the PNCS must be reported in the same way as if they fell within the framework of the PNCS.

When Salmonella spp is isolated in samples taken in the course of operator own checks, the laboratories shall serotype them in order to be able to distinguish at least between the serotypes covered by this programme and other Salmonella spp serotypes. The laboratory may carry out the serotyping itself or send the samples to another laboratory authorised under the PNCS in accordance with point 12 of this Programme for serotyping. If the serotyping shows positive for one of the serotypes in question or for any other serotype, or if their presence cannot be ruled out, and the initial sample was taken in an own check, it must be reported to the competent authority as soon as possible, and never later than 24 hours after the laboratory or the operator of the holding operator receives the results of the analysis.
As soon as the operator becomes aware of the existence of a positive result he shall be responsible for taking the appropriate measures, as set out in this programme for cases where any of the Salmonella serotypes covered by the programme are detected. The competent authority may exceptionally carry out a confirmatory analysis if it considers this appropriate.
All the results of own checks must be recorded using the dedicated computer application used by the authorised laboratories to communicate results, without prejudice to the contents of the previous paragraph. To ensure suitable traceability of the samples taken during own checks and official monitoring and in order to ensure suitable computer processing of the sampling data for this programme, the sampled flocks shall be identified as specified in Point 3 of this Programme.

The competent livestock service and health authorities must keep each other suitably informed of the positive results.

5. Biosecurity measures

FBOs have to implement measures to prevent the contamination of their flocks.

☒*yes* ☐*no*

If no please explain. if yes,describe also the biosecurity measures that shall be applied, quote the document describing them (if any) and attach a copy

Biosecurity measures will be verified in accordance with the protocol included in this programme for checking biosecurity measures on breeding poultry holdings.

These checks will take place in the course of each of the official inspections provided for on the holdings, at the frequency indicated in this Programme. The data gathered in such surveys must be recorded using the MAPA computer application for official inspections, in the 'biosecurity' section.

If, in the course of an inspection, shortcomings in the biosecurity measures are detected, this will be made known to the owner of the holding by means of a report in at least triplicate for the owner of the holding and his legal representative or the person in charge of the animals, setting out all the shortcomings and the deadlines set for them to be remedied.

The veterinary officer shall adopt a proportionate and progressive approach in his work to enforce biosecurity rules and measures.
The competent authority may, if necessary, make use of the measures established in Chapter IV, Title V, of Law 8/2003, the Animal Health Act. This is without prejudice to other measures or penalties which may be adopted in respect of that flock or throughout the holding, depending on the type of shortcoming. The measures to be adopted to prevent health risks depend on the seriousness of the shortcoming and may range from shutting down the holding to the loss of the health authorisation for operating a holding.
The attached procedure will be followed to check and improve biosecurity measures in breeding poultry holdings.

6. Minimum sampling requirements for food business operators :

Samples at the initiative of the FBOs will be taken and analysed to test for the target *Salmonella* serovars respecting the following minimum sampling requirements:

a. Rearing flocks: day-old chicks, four-week-old birds, two weeks before moving to laying phase or laying unit

b. Adults breeding flocks: depending if the MS achieved the EU target for more than 2 years

Breeding flocks of Gallus gallus

☐ Every second week during the laying period (at the holding and at the hatchery)

☒ Every three weeks during the laying period at the holding. Sampling frequency remains at every 2nd week at the hatchery. (derogation of point 2.1.1 of Annex to Regulation (EC) No 200/2010)

Comments - Indicate also who takes the FBO samples

Sampling shall be carried out in accordance with the minimum requirements laid down in Part B of Annex II to Regulation (EC) No 2160/2003 of the European Parliament and of the Council. Zoonosis / Zoonotic agent Salmonella spp with public health significance (ST, SE, SH, SV, SI)
Stages of production to be covered by sampling
Rearing:
I. day-old chicks
II. 4-week-old birds
III. two weeks before transfer to the laying unit or the start of the laying phase
Adults:
II. Every 2 weeks during the laying phase

The owner of the holding shall be responsible for carrying out own checks, including sampling, in the form and under the conditions provided for by this programme. Sampling may be carried out by qualified staff from the laboratory which performs the analyses. The veterinarian responsible for the holding will ensure that the sampling protocol is in accordance with the conditions laid down in this programme.
Since the Community target has been reached at national level for at least two consecutive calendar years in Spain, the frequency of sampling on the holding may be extended to every three weeks, at the discretion of the competent authority and in accordance with Commission Regulation (EC) 213/2009. Each Autonomous Community is responsible for authorising the extension of the frequency of sampling in its territory.
The owner of the holding shall keep the results of the analysis for a period of at least three years, during which time they will be at the disposal of the competent authority. Recording of results in the Ministry own-check application.
The data and information obtained from holdings where official sampling is performed (Annex: OWN-CHECK sampling) and the laboratory results shall be recorded in the application of the National programme for the control of Salmonella https://servicio.mapa.gob.es/ The results of the own-check samples must be recorded in the own-check application, together with the required accompanying data, within one month of the laboratory analysis result being obtained; the results must be obtained within 10-15 days of the sampling, on average, except in exceptional circumstances. All of the data from the sampling sheet must be filled in correctly: if any information is missing the samples cannot be recorded in the application. All samples and data relating to sampled flocks that are not recorded in the Ministry applications (official monitoring and own checks) will not be valid within the framework of the PNCS.
The above notwithstanding, all positive results for Salmonella considered to have public health significance must be notified as specified in the PNCS.

7. Samples are taken in accordance with provisions of point 2.2 of Annex to Regulation (EU) No 200/2010

☒*yes* ☐*no*

If no please explain.

A. MINIMUM SAMPLING REQUIREMENTS FOR OWN CHECKS
Sampling must observe the minimum sampling requirements laid down in Part B of ANNEX II to Regulation (EC) No 2160/2003 of the European Parliament and of the Council and in the ANNEX to Commission Regulation (EU) No 200/2010 of 10 March 2010 implementing Regulation (EC) No 2160/2003 of the European Parliament and of the Council as regards a Union target for the reduction of the prevalence of Salmonella serotypes in adult breeding flocks, and Commission Regulation (EC) No 213/2009 amending Regulation (EC) No 2160/2003 and Commission Regulation (EU) 2019/268 of 15 February 2019 amending Regulations (EU) No 200/2010, (EU) No 517/2011, (EU) No 200/2012 and (EU) No 1190/2012 as regards certain methods for Salmonella testing and sampling in poultry.

A.1. Sampling in adult breeding flocks (both own checks and official controls)
Sampling will involve obtaining sufficient faecal samples to detect 1% of infected birds in the flock with a 95% confidence limit. To that effect, the samples shall comprise one of the following:
a) Pooled faeces obtained from individual samples of fresh faeces weighing not less than 1 g, taken at random from various parts of the building in which the poultry are kept, or where the birds have free access to more than one building on a particular holding, from each group of buildings to which the flock has access. The faeces shall be pooled and a minimum of 2 pooled samples per flock analysed. The number of sites from which separate faeces samples are to be taken in order to make a pooled sample shall be as follows:

Number of birds in the breeding flock	Number of samples of faeces to be taken from the breeding flock
250-349	200
350-449	220
450-799	250
800-999	260
1000 or more	300

b) Boot swab samples, comprising 5 pairs of absorbent boot swabs. The laboratory will handle the boot swabs as 2 composite samples, each one comprising 5 boot swabs. Boot swabs used shall be sufficiently absorptive to soak up moisture. Tubegauze 'socks' shall also be acceptable for that purpose. The surface of the boot swab shall be moistened using appropriate diluents (such as 0.8 % sodium chloride, 0.1 % peptone in sterile deionised water, sterile water or any other diluent approved by the competent authority). Furthermore, measures shall be taken to prevent the potential bacterial growth inhibiting effects of the disinfectants used in the foot baths at the entrances to the sheds. The samples shall be taken while walking through the house using a route that produces representative samples for all parts of the poultry house or the respective sector. It shall include littered and slatted areas provided that slats are safe to walk on. All separate pens within a poultry house shall be included in the sampling. On completion of the sampling in the chosen sector, boot swabs must be removed carefully so as not to dislodge adherent material. The boot swabs shall be placed in a bag, flask or other type of sterile container which shall then be sealed and labelled appropriately.
c) For caged flocks, sampling shall consist of naturally mixed faeces from dropping belts, scrapers or deep pits, depending on each holding's dropping collection system. Two samples of at least 150 g each

shall be collected to be tested individually. As there are normally several stacks of cages within a house and all must be represented in the sample, the sample shall be taken as described below: -In systems where there are belts or scrapers, these shall be run on the day of the sampling before sampling is carried out in order to collect only fresh faeces. -In systems where there are deflectors beneath cages and scrapers, droppings which have lodged on the scraper after it has been run shall be collected. -In systems where the droppings empty directly into a pit, the droppings shall be collected directly from the pit.

(d) In cage houses where a sufficient amount of faeces does not accumulate on scrapers or belt cleaners at the discharge end of belts, four or more moistened fabric swabs of at least 900 cm2 per swab, moistened using appropriate diluents (such as 0,8 % sodium chloride, 0,1 % peptone in sterile deionised water, sterile water or any other diluent approved by the competent authority, shall be used to swab as large a surface area as possible at the discharge end of all accessible belts after they have been run, ensuring each swab is coated on both sides with faecal material from the belts and scrapers or belt cleaners.
(e) In multi-tier barn or free range houses in which most of the faecal material is removed from the house by dropping belts, one pair of boot swabs shall be taken by walking around in littered areas in accordance with point (b) and at least 2 moistened fabric swabs shall be taken as hand-held swabs from all accessible dropping belts, as in point (d).

A.2. Sampling in rearing flocks
The following procedure will be adopted in rearing flocks:

a) Day-old chicks:
1. One sample made up of from 10 samples taken of the internal coverings of the cages transporting the chicks taken when they are delivered to the holding. The bases of the cages may be used directly as a sample, which will be sent either whole or in parts to the laboratories responsible for processing samples and may be made up of a single or more than one sample, or
2. Liver, caecum and yolk sac of 60 chicks (these parts of the viscera can be removed and processed as a single sample), or
3. A sample made up of meconium from at least 250 chicks.

b) 4-week-old birds, and birds two weeks before transfer to the laying unit (or the start of the laying phase):
1. A sample of portions of fresh faeces of a minimum weight of one gram each collected at random at a minimum of ten different points in accordance with the following table: Faeces may be pooled for analysis up to a minimum of two pools.
Number of birds kept in one house // Number of portions of faeces to be taken in the house/group of houses on the holding
1-24 (equivalent to the number of birds up to a maximum of 20)
25-29 20
30-39 25
40-49 30
50-59 35
60-89 40
90-199 50
200-499 55
500 or more 60
2. The samples shall comprise 5 pairs of absorbent boot swabs. The laboratory will handle the boot

swabs as 2 composite samples, each one comprising 5 boot swabs.

Preparation of the samples in the laboratory (official controls and FBO controls):

a) Boot swabs and fabric swabs:
The pair(s) of boot swabs must be unpacked carefully to avoid dislodging adherent faecal material. They must be collated into two samples and submerged in 225 ml buffered peptone water (BPW) that has been pre-warmed to room temperature. If necessary, more peptone water may be added to leave liquid around the sample to permit migration of Salmonella. Shake to ensure complete saturation of the sample and continue to apply the detection method.
In case of collection of fabric swabs in accordance with point 7.A.1(d) and e) of this programme pooling shall occur fully submersing boot/socks and fabric swab in BPW to provide sufficient free liquid around the sample for migration of Salmonella away from the sample and therefore more BPW may be added, if necessary.
Separate preparations must be made of the boot swabs and the fabric swab.

b) Other faeces samples and dust samples: - The faeces samples shall be pooled and thoroughly mixed for analysis into a minimum of two pools and a 25-gram sub-sample shall be collected from each one for the culture. - Add 225 ml buffered peptone water to the 25-g sub-sample and shake gently. - Culturing of the sample shall be continued by using the detection method set out in point C. For preparation of all of these samples, Standard UNE-EN ISO 6887-6, “Specific rules for the preparation of samples taken at the primary production stage”, may also be used as a guide.

Identification of samples and results of analyses (official controls and own checks):

The samples sent must be properly preserved and identified (in accordance with the specimen report accompanying the samples to the laboratory, included in the annexed Sampling Sheet). There are two standard sampling sheets: one for official controls and one for own checks, since it is not necessary to collect as much information for own checks as for official controls. In both cases it must be clearly indicated that the samples are taken within the framework of the PNCS to avoid any confusion with private samples taken by the holding. The sampling sheets are to be completed in their entirety since all of the information collected on the forms is required for assessment of the PNCS. One copy or a duplicate of the sampling sheet must remain on the holding and must be filled with the test report sent by the laboratory so that all of the documentation relating to the samples is present on the holding (sampling sheet and test results). This documentation must be available to the official veterinary services when official controls are carried out in the framework of the PNCS . The documentation required may be submitted in hard copy or electronic format.
To ensure suitable traceability of the samples, at least the following information must be recorded in the test reports:
1. Date on which samples were taken.
2. Identification of the flock. (REGA, CAPITAL LETTER IDENTIFYING THE SHED, DATE OF ENTRY OF THE BIRDS INTO THE SHED (format mmyyyy). 3. Poultry population (breeders, layers, broilers, fattening or breeding turkeys) 4. Samples (specimen, number and weight or volume) that arrived at the laboratory and method by which they were mixed for analysis. All reports on tests carried out on samples as part of the PNCS, and the annexed sampling sheets, must include the following text, clearly and easily visible: “THESE SAMPLES FALL UNDER THE SALMONELLA NATIONAL CONTROL PROGRAMMES”.

When a vaccine strain has been detected, the laboratory serotyping report must include the following statement : " The flock shall be considered negative because it has been isolated a vaccine strain"

(this text was added after a request of the FBO in order to clarify the status of the flock and to avoid trade misunderstanding).

8. Specific requirements laid down in Annex II.C of Regulation (EC) No 2160/2003 will be complied with where relevant (i.e. due to the presence of SE or ST (including monophasic ST 1,4,[5],12:i:-), all birds of infected rearing or adult flocks are slaughtered or killed and destroyed, and all eggs are destroyed or heat treated):

☒*yes* ☐*no*

If no please explain. Indicate also if birds are slaughtered or killed and destroyed, and if eggs are destroyed or heat treated.

The minimum measures to be adopted when the presence of S. Enteritidis, S. Typhimurium, including the monophasic variant of Salmonella Typhimurium with the antigenic formula 1,4,[5],12:i:-, S. Hadar, S. Virchow and/or S. Infantis is detected in a flock of birds are as follows:
1. An in-depth epidemiological investigation shall be carried out to attempt to identify the cause of the positive result and detect the source of infection, in accordance with the epidemiological survey attached in the programme. Where appropriate, official samples may be taken of the feed and/or water used on the holding or given to the positive flock.
2. No live birds may be moved into or out off this site unless prior authorisation has been obtained for them to leave for the purposes of slaughter or destruction. Any transfer of animals must be accompanied by a health document made out by the competent authority stating at least the number of animals and the necessary information for identifying the holding and the transporter.
3. All birds, including day-old chicks, in the flock must be slaughtered or destroyed so as to reduce as much as possible the risk of spreading salmonella. Slaughter must be carried out in accordance with Community legislation on food hygiene. Products obtained from these birds may be placed on the market for human consumption only in compliance with Community legislation on food hygiene in force and with part E of Annex II to Regulation (EC) No 2160/2003. If not destined for human consumption, such products must be used or disposed of in accordance with Regulation (EC) No 1069/2009 of the European Parliament and of the Council of 21 October 2009 laying down health rules concerning animal by-products not intended for human consumption and repealing Regulation (EC) No 1774/2002.
4. Non-incubated eggs from the flock must be destroyed. However, such eggs may be used for human consumption if they are treated in a manner that guarantees the destruction of Salmonella in accordance with Community legislation on food hygiene and with the provisions of part D of Annex II to Regulation 2160/2003.
5. Where eggs for hatching from flocks in which one of the five serotypes of Salmonella has been confirmed are still present in a hatchery, they must be destroyed or treated in accordance with Regulation (EC) No 1069/2009.
6. Thorough checking of biosecurity measures for all flocks on the holding in accordance with the procedure for checking biosecurity measures on breeding poultry holdings.
7. Once the birds from the infected flock have been slaughtered or destroyed, efficient and thorough cleaning (including complete removal of the bedding and excrement) shall be undertaken, followed by disinfection, insect removal and rat extermination. The above tasks shall be performed using properly authorised and registered products. As soon as sufficient time has elapsed after disinfection, environmental samples shall be taken to check the effectiveness of the cleaning and disinfection process

and the absence of Salmonella spp. in the environment. Verification of cleaning and disinfection should be done according to point 17 of the programme, which describes that the competent authorities shall check the suitability of the cleaning, disinfection and depopulation measures adopted in the hen houses and where appropriate, will authorise installations to be occupied by new animals. For the cleaning and disinfection procedure to be considered valid, a minimum of 10 samples (dust, fabric swabs, chamois or similar sampling materials) must be taken at various points on the holding and must yield negative results for Salmonella spp. Samples may be combined to produce a single culture. The use of cotton swabs or brushes is not recommended as they only pick up very small quantities of sample.
8. The premises shall not be restocked for 12 days after completion of the cleaning, disinfection, rat extermination and if necessary insect removal processes. Repopulation may take place only if the environmental analyses carried out in accordance with the programme are satisfactory, and if biosecurity measures considered inadequate or deficient by the competent authority have been properly corrected. Notwithstanding the above, in those cases where the results of those tests prove the effectiveness of the cleaning and disinfection undertaken, the waiting period may be reduced to a minimum of 7 days.
9. The dates of slaughter or destruction of the flock, disinfection, collection of environmental samples and restocking must be notified to the competent authorities. All of these processes must be duly recorded for possible consultation by the competent authorities and any depopulation, slaughter or destruction of the flock and restocking must take place under official supervision.
10. Where one of the five types of Salmonella is confirmed on heavy breeder holdings, the above-mentioned measures at least shall be adopted and, in addition, the next batch of birds introduced must be pullets vaccinated with authorised vaccines or autovaccines in accordance with the legislation in force, before commencing the laying stage.
11. If necessary, it may be requested the results of laboratory analyses of the worker/s in charge of the animals or anybody who can be considered as a risk in order to determine whether there are any Salmonella spp. carriers among them. (This text was requested by the sector)

If, however, a serotype not concerned by the control programme is identified, the following measures will be taken:
1. An in-depth epidemiological investigation shall be carried out to attempt to identify the cause of the positive result and to detect the source of infection. Where appropriate, official samples may be taken of the feed and/or water used on the holding or given to the positive flock.
2. Thorough checking of biosecurity measures for all flocks on the holding in accordance with the procedure for checking biosecurity measures on breeding hen holdings.

9. Please describe the measures that shall be implemented in a flock (rearing and adult) where ***Salmonella*** **Hadar, Infantis or Virchow is detected**:

(max. 32000 chars) :

Exactly the same measures must be taken as when S. Enteritidis or S. Typhimurium, including the monophasic variant with the antigenic formula 1,4,[5],12:i:-, are detected.
These measures are described in Section 8.

10. If birds from flocks infected with SE or ST are slaughtered, please describe the measures that shall be implemented by the FBO and the CA to ensure that fresh poultry meat meet the relevant **EU microbiological criteria** (row 1.28 of Chapter 1 of Annex I to Regulation (EC) No 2073/2005): absence of SE/ST in 5 samples of 25g:

Measures implemented by the FBO (farm level)

++ (In order to clarify the SNCP of poultry, this text is amended as a part of the Action Plan aproved after the recommendation of report ref DSG(SANTE) 2019-6597 of the EU audit to evaluate SNCP carried out in November 2019, stating that the CA should ensure that only broiler and turkey flocks that have been sampled for Salmonella with a known test result can be sent for slaughter)++
In accordance with Royal Decree 361/2009 on food chain information, the operator of the livestock holding must ensure that in all shipments of animals to the slaughterhouse, full information on the results of all analyses of samples taken that have importance for human health, in the framework of the surveillance and control of Salmonella is sent to the slaugtherhouse operator; in other words, the slaughterhouse operator must be informed if the result of the last analysis (or the results of last analyses, if several samples have been taken in the near future) has been negative or positive to Salmonella spp. and, in this last case, in addition, if it is negative or positive to S. Enteritidis or S. Typhimurium, and the information of the result/s of such analysis must be included in the FCI (Food Chain Information) to be considered complete.
If a flock on the holding tests positive for S. Enteritidis or S. Typhimurium, the operator of the livestock holding must also ensure that no live birds are moved into or out off this site unless prior authorisation has been obtained for them to leave for the purposes of slaughter or destruction. Any transfer of animals must be accompanied by a health document to be drawn up and completed by the competent authority indicating at least the number of animals and the information necessary to identify the holding and the transporter.

Measures implemented by the FBO (slaughterhouse level)

Slaughter at the slaughterhouse shall be carried out in accordance with the provisions of Regulation (EC) No. 853/2004, which lays down specific hygiene rules for food of animal origin, and in particular Section II of Annex III thereof.
When a positive herd is received at the slaughterhouse, it is logistically slaughtered, i.e. the herd is slaughtered last in the daily slaughter order to minimize the possibility of cross-contamination, followed by cleaning and disinfection. This is carried out in line with the provisions of Regulation (EU) 2019/627 with the aim of reducing contamination of other animals or their meat as much as possible. In addition, in accordance with the provisions of Regulation (EC) No. 2073/2005, slaughterhouses shall include in their sampling plans poultry carcasses from flocks whose Salmonella status is unknown or positive for Salmonella Enteritidis or Salmonella Typhimurium.
There is a “Manual for the broiler sector in Spain for compliance with Regulation (EU) No 1086/2011 amending Regulations (EU) No 2160/2003 and (EC) No 2073/2005”, which, although it is voluntary, can provide guidance as to the correct way of handling birds slaughtered in slaughterhouses in relation to Salmonella.
As an example of the possible system of action, attached is the management diagram of birds sent to a slaughterhouse, recommended in the "GUIDE FOR THE MEAT POULTRY SECTOR IN SPAIN FOR COMPLIANCE WITH REGULATION (EU) No. 1086/2011 AMENDING REGULATIONS (EU) No. 2160/2003 AND (EC) No. 2073/2005", with some additional issues that are carried out voluntarily by the

slaughterhouses that apply the guide, such as the immobilization of the carcasses sampled until the results are available.
Guide available through: https://www.aesan.gob.es/AECOSAN/docs/documentos/seguridad_alimentaria/gestion_riesgos/PROPOLLO.pdf

Measures implemented by the CA (farm and slaughterhouse level)

The official veterinarian is responsible for verifying that the correct food chain information is passed on as required pursuant to RD 361/2009: accordingly, he or she must check that the livestock holdings are passing this information to the slaughterhouses in a consistent and effective, valid and reliable manner and ensure that the relevant animal health and food safety information, including that relating to the results of Salmonella testing, is also passed on. Provision is thus made for slaughterhouses to only accept animals for which the relevant information on the holding of origin has been received. As a general rule the information should be received at least 24 hours prior to the arrival of the animals. Slaughter in slaughterhouses must take place in accordance with Regulation (EC) No 853/2004 laying down specific hygiene rules for food of animal origin, and in particular Section II of Annex III.

Official controls must be carried out in accordance with Commission Delegated Regulation (EU) 2019/624 of 8 February 2019 concerning specific rules regarding the performance of official controls on meat production and regarding production and relaying areas for live bivalve molluscs in accordance with Regulation (EU) 2017/625 of the European Parliament and of the Council and Commission Implementing Regulation (EU) 2019/627, of 15 March 2019 laying down uniform practical arrangements for the performance of official controls on products of animal origin intended for human consumption in accordance with Regulation (EU) 2017/625 of the European Parliament and of the Council and amending Commission Regulation (EC) No. 2074/2005 of the European Parliament and of the Council and Commission Regulation (EC) No. 2074/2005 of the European Parliament and of the Council. No. 2074/2005 of the Commission as regards official controls.

The provisions of Regulation (EC) No 2073/2005, on microbiological criteria for foodstuffs, also apply in relation to the criteria for Salmonella in poultry meat. Once positive results for S. Enteritidis or S. Typhimurium are found in a consignment, the official veterinarian will ensure that targeted sampling and tests using the EN/ISO 6579 methodology or a validated alternative method are carried out, and lastly that the carcasses are withdrawn from the market and destroyed or that the destination previously given for the product is changed.

11. Laboratories in which samples (official and FBO samples) collected within this programme are analysed are accredited to ISO 17025 standard and the analytical methods for *Salmonella* detection is within the scope of their accreditation.

☒*yes* ☐*no*

If no please explain.

The Central Veterinary Laboratory in Algete (Madrid) of the Ministry of Agriculture, Fish and Food is the National Reference Laboratory for all serotypes of Salmonella in animals.
Laboratories analysing official samples as part of the programme must be established, recognised or designated by the competent bodies in the Autonomous Communities. These official laboratories must operate and have access to accredited tests for Salmonella in all matrices monitored under the PNCS

with which they work, and be accredited in accordance with Standard EN/ISO 17025 on general requirements for the competence of testing and calibration laboratories, or must apply quality assurance systems in accordance with that standard. They must also participate in the ring tests organised or co-ordinated by the National Reference Laboratory.
The laboratories participating in the programme for the purposes of carrying out own checks must be recognised by the competent authorities of the Autonomous Communities in which they are established and must operate and have access to accredited tests for Salmonella in all matrices monitored under the PNCS with which they work, and be accredited in accordance with Standard EN/ISO 17025 on general requirements for the competence of testing and calibration laboratories, or apply quality assurance systems in accordance with that standard. Laboratories must also regularly participate in collaborative testing organised or co-ordinated by the National Reference Laboratory.
The list of participating laboratories must be published, for information purposes, at least on the MAPA website.
The competent authorities of the Autonomous Communities shall notify the Ministry of Agriculture, Fish and Food of the laboratories referred to in the previous paragraph or of any modifications to them so that the list may be published at least on the departmental website for information purposes.
Where a laboratory serves at the same time as an Autonomous Community's official laboratory and participates in the own-check programme, it must notify the relevant competent authority or authorities and ensure that the two activities are managed separately, and is subject to monitoring and periodic inspection by the competent authority to check that these are separate. If it fails to notify the authorities, or cannot guarantee that the activities are kept separate, it cannot operate as an official laboratory.
The results obtained by authorised laboratories for both official monitoring and own checks shall be valid and applicable throughout the country.
Laboratories must reject samples which do not meet the requirements specified in this programme.

12. The **analytical methods** used for the detection of the target *Salmonella* serovars is the one defined in Part 3.2 of the Annex of Regulation (EU) No 200/2010 i.e. Amendment 1 of EN/ISO 6579-2002/Amd1:2007. `*Microbiology of food and animal feeding stuffs - Horizontal method for the detection of Salmonella spp. -- Amendment 1: Annex D: Detection of Salmonella spp. in animal faeces and in environmental samples from the primary production stage*'.

    Serotyping is performed following the Kaufman-White-Le Minor scheme.

☒*yes* ☐*no*

If yes, please describe the alternative method(s) used.

Salmonella spp. shall be isolated in accordance with Standard EN/ISO 6579 -1 (update to Regulation (EU) 2019/268). Horizontal method for the detection of Salmonella spp. in animal faeces and in samples at primary production level" which uses a semi-solid culture medium (modified semi-solid Rappaport - Vassiladis - MSRV) as a single selective enrichment medium. The semi-solid medium should be incubated at 41.5 ± 1 °C for 2x (24±3) hours. At least one isolate from each sample showing a positive reaction shall be typed, in accordance with the Kaufmann-White-Le Minor scheme. Laboratories may type their own Salmonella isolates or send them to other laboratories authorised within the PNCS to be typed. The laboratory where typing takes place must issue a report including its results and send it to the laboratory

that sent the isolates to be typed. The recording of results in the application and the notification of results as indicated in this programme are the responsibility of the laboratory that isolated the Salmonella. To prevent any delays in obtaining and notifying the results of typing:
• The isolate must be sent to another laboratory for typing no more than 24 hours following isolation.
• Typing must begin in the laboratory no more than 24 hours following receipt of the isolate in the laboratory.
• The issue and dispatch of the results report from the typing laboratory to the laboratory that sent the isolate, or the notification of the results, as appropriate, must take place no more than 24 hours after the results are obtained in the laboratory.
• The recording in the application and the notification of positive results by the isolating laboratory must take place within the deadlines laid down in this programme.

For samples taken on behalf of the FBO alternative methods if validated in accordance with the most recent version of EN/ISO16140 may be used.

☒*yes* ☐*no*

If no please explain. If timelimits are exceeded, please indicate what is done.

Alternative methods may be used instead of the methods referred to above, if validated in accordance with EN ISO 16140-2 (for alternative detection methods). (Update to Regulation (EU)2019/268).

13. Samples are transported and stored in accordance with point 3.1.1 of the Annex to Regulation (EU) No 200/2010. In particular, samples examination shall start in the laboratory within 48 hours following receipt and within 96 hours after sampling.

☒*yes* ☐*no*

If no please explain.

Samples shall be sent to the laboratories referred to in Articles 11 and 12 of Regulation (EC) No 2160/2003, within 24 hours after collection, preferably by express mail or courier. If not sent within 24 hours, they must be stored refrigerated. They may be transported at ambient temperature as long as excessive heat (over 25°C) and exposure to sunlight are avoided. At the laboratory samples shall be kept refrigerated until examination, which shall be started within 48 hours of receipt and within 96 hours of sampling.

14. Please describe the **official controls at feed level** (including sampling).

Comments (max. 32000 chars) :

Control measures to prevent the introduction of Salmonella spp. in farms through feed are based on the verification of compliance with current feed regulations by the competent authority of the Autonomous Regions.
As described in Article 15 of Regulation (EC) 178/2002 laying down the general principles and requirements of food law, establishing the European Food Safety Authority and laying down procedures in matters of food safety, the feed operator shall not place unsafe feed on the market which has an adverse effect on human or animal health or which renders the feed obtained from food-producing animals unsafe for human consumption. Therefore, the operator shall take necessary, effective, proportionate and specific measures to continuously minimize potential Salmonella contamination and protect human and animal health.The producer of the feed material shall establish, implement and maintain a permanent written procedure or procedures based on HACCP principles in accordance with Article 6 of Regulation (EC) 183/2005 laying down requirements for feed hygiene. Procedures based on HACCP or guidelines are aimed at significantly reducing the presence of Salmonella and minimizing the re-contamination of the final product or reducing the level of contamination, according to the specific risk assessment of each operator through a strict system of controls throughout the process and the application of various measures aimed at reducing the risk of Salmonella spp. presence.The critical points of the manufacturing process will depend on each operator and will have to take into account the evaluation and control of suppliers (microbiological quality of the raw materials supplied or other factors that may compromise it), the application of cleaning programs and the application of good practice guidelines throughout the production chain (storage of raw materials, manufacturing, storage of the finished product, etc.).
The control measures by the competent authority of the Autonomous Regions include different aspects such as the verification of the purchase of feed from registered or authorized operators, in accordance with Regulation (EC) 183/2005 laying down requirements for feed hygiene, including the application of systems and self-monitoring based on HACCP principles and guides to good hygiene practices. The objective is to ensure that no Salmonella contamination occurs during the processing of poultry feed, guaranteeing feed safety at all stages that may have an impact on feed and food safety, including the primary production of feed and food.It should also be noted that Regulation (EC) No. 183/2005 on Feed Hygiene, applicable since January 1, 2006, requires the establishment of harmonized microbiological criteria, based on scientific criteria of Risk Analysis, to harmonize intra-Community trade and ensure that imported feed complies with levels at least equivalent to those produced in the national territory. According to this Regulation, feed exporting companies must comply with specific microbiological criteria. The criteria and targets must be adopted by the EU in accordance with the procedure laid down in Article 31 of the Regulation. Feed business operators responsible for the primary production of feed must take the necessary measures to prevent, eliminate or reduce feed safety risks during the procurement and storage of raw materials and the subsequent stages of manufacture, preparation, cleaning, packaging, storage and transport of such products (as referred to in Annex I of Regulation 183/2005). They must also keep records detailing the measures taken to control contamination hazards. Other feed business operators must take appropriate measures to ensure the safety of the products they manufacture, transport or use. These measures are more precisely detailed in Annex II of the aforementioned regulation, and they shall apply the principles of the HACCP system, taking corrective measures when the monitoring of a critical point is not controlled and implementing internal

procedures to verify that the measures taken are effective. They must also maintain records in order to demonstrate the application of these measures.
Therefore, feed hygiene requirements are verified in all the activities of operators in the animal feed sector, from the primary production of feed to its commercialization, as well as the feeding of food-producing animals and the import and export of feed from and to third countries, with the purpose of adopting the appropriate measures to guarantee the safety of feed at each stage.
It should be noted that there is no Community or national regulation establishing microbiological criteria for Salmonella (or other microorganisms) in raw materials and feed of vegetable origin, although there are legal criteria established for raw materials and feed of animal origin.
The program of official controls in animal feed, approved within the National Coordination Commission for Animal Feed (CNCAA), indicates that, given that, in the case of vegetable products (whether raw materials or feed), these determinations do not have a maximum limit established in the current national or Community regulations, in the event of a positive result for Salmonella, an identification of the serotype must be requested. Only in the case of S. Enteritidis, S. Typhimurium, S. Infantis, S. Virchow and S. Hadar, notification will be made through the Alert Network.
In case of a positive result for Salmonella spp, the approved HACCP system must apply corrective measures that allow the product, in a new analytical control, to demonstrate that it is suitable to be placed on the market. These measures are included in international, community and national sectoral guides. This is the case of the Guide for the development of feed sanitization standards, prepared in 2007 by the Spanish Confederation of Compound Feed Manufacturers (CESFAC), which compiles in a single document the possible sanitization systems that can be applied in a factory to obtain microbiologically safe feed, such as heat treatment or the use of authorized additives.Available at: https://cesfac.es/media/attachments/2019/08/08/guia-higienizacin.pdf
The information on the authorization of feed additives, contained in the guides, must be verified with the register of authorized additives which can be accessed through the following link: https://food.ec.europa.eu/safety/animal-feed/feed-additives/eu-register_en
There are no criteria to be followed in the EU zoonosis regulations regarding the potential presence of Salmonella and other potential zoonotic agents in feed. The sampling that accompanies the official controls on establishments that destine products for animal feed includes analytical determinations to detect the presence of Salmonella in raw materials and feed. In the case of products of plant origin, analytical determinations are carried out taking into account the risk criteria established in public documents approved by the CNCAA in which possible hazards to be controlled in raw materials intended for the manufacture of animal feed and, therefore, in the feed of which they are part (DOC CNCAA 1/2015 vers 1. Main hazards to be controlled in self-control systems). This document has been disseminated to operators in the sector through their associations, the control authority, and is accessible on the SILUM application on the website of the Ministry of Agriculture, Fisheries and Food: https://www.mapa.gob.es/es/ganaderia/temas/alimentacion-animal/acceso-publico/pruebaotros.aspx
Every year, more than 3,000 official inspections are carried out in national establishments destined for animal feed products, verifying the self-controls performed by operators in the sector and more than 1,000 official samples are taken for the determination of microbiology, including Salmonella. These data are included in the PNCOCA annual report, distributing the samples among raw materials, compound feed and other products.

15. Official controls at holding, flock and hatchery level

a. Please describe the official checks concerning the **general hygiene provisions** (Annex I of Regulation (EC) No 852/2004) including checks on biosecurity measures, and consequences in case of unsatisfactory outcome.

(max. 32000 chars) :

"Guides to Good Hygiene Practice for the prevention of zoonotic Salmonella in holdings for the selection, breeding and rearing of flocks of Gallus gallus" have been drawn up jointly by representatives of the breeding poultry sector and the Ministry of Agriculture, Food and the Environment.
They are available in printed form for distribution to livestock farmers in the sector and the competent authorities.
They are also available for consultation on MAPA's website: http://www.mapa.es/.
Holders of breeding hen establishments must have in place a code of good hygiene practices in order to meet the objective of this national Salmonella control programme and to ensure that health information is kept up-to-date. They must also keep the following records on holdings:
a) A record of the type and origin of the feed supplied to the animals.
b) A record of the outbreak of diseases that could affect the safety of products of animal origin.
c) An up-to-date record of visits, listing the people and vehicles that have entered the holding.
d) A record of medicinal treatments, containing the information specified under Article 8 of Royal Decree 1749/1998 setting out the applicable control measures for certain substances and their residues in live animals and their products and including the vaccinations referred to in this programme.
e) All the results of analyses and checks to detect Salmonella carried out on the flock concerned, including those carried out in the incubator or breeding shed of origin of the flock, must be kept by the owner of the holding for at least three years and the records of the flock currently in production must, without fail, be kept on the holding.
f) All movements of flocks entering and leaving the holding must be recorded in the holding register. The flock sheet must be kept for at least three years after the flock is slaughtered.
g) There must also be a documentary record of:
i. The protocols and records of cleaning and disinfection work (dates, products used, the person or company responsible for this work).
ii. Analyses to check that cleaning and disinfection operations carried out during the depopulation period have been effective in guaranteeing control of Salmonella with public health significance.
iii. Rat and insect extermination programmes and implementation records (dates, products used, procedure for verifying the effectiveness of the programme, etc.)
h) Producers of rearing pullets must report on the health status of the breeding flock of origin and on any vaccinations and own checks during the rearing of the pullets; this information must accompany the pullets when they are transferred to the producing holdings.
The owner of the holding must be in possession of all the mandatory health documentation and keep records of all of the necessary data so that the competent authority can regularly check compliance with the health programme referred to in this paragraph as well as the code of good hygiene practices, in particular the records mentioned above (a), b), c), d) and e)).

Without prejudice to Royal Decree 637/2021, the holder must adopt protective livestock rearing measures to control the introduction of or contamination by Salmonella spp on the holding. In

particular:
a) The design and maintenance of the installations must be suitable for preventing the entry of Salmonella spp.;
b) Appropriate measures must be taken to control rodents, insects, wild birds and other domestic or wild animals which might introduce the disease. A rat extermination programme must be carried out either by the holding itself or by authorised establishments.
c) Day-old chicks must be obtained from holdings and hatcheries which have satisfactorily passed inspections to prevent the vertical transmission of the five Salmonella serotypes; the supplier must certify that the said chicks are exempt from the five abovementioned serotypes, and documentary evidence of the favourable outcome of laboratory tests must be made available to the purchaser. Rearing pullets (future layers) must be accompanied when leaving the rearing establishment by a certificate from the supplier stating that own checks have been properly carried out and detailing their results (day-old chicks and birds two weeks before entering the laying stage or unit must have satisfactorily passed the tests for the five Salmonella serotypes). Where appropriate, they shall also be accompanied by a certificate stating that the pullets have been vaccinated in accordance with the programme. These requirements must be met before authorisation is given for the transfer and restocking of the laying shed.
d) Adequate washing, cleaning, disinfection and rat extermination measures must be taken in rearing houses, breeding hen houses and adjoining structures and also with regard to the material and equipment used for productive activity.
e) Tests must be conducted to verify that cleaning and disinfection were carried out correctly. To verify cleaning and disinfection two or more moistened fabric swabs of at least 900 cm2 per swab, moistened using appropriate diluents (such as 0,8 % sodium chloride, 0,1 % peptone in sterile deionised water, sterile water or any other diluent approved by the competent authority), shall be used to swab as large a surface area in different points in the house (floor, walls, feeding equipments, watering equipments, belts, pilars, water and feeding pipes, scrapers and any other difficult point to clean and disinfect). Samples can be pooled to perform and single culture, or by enriching the peptone water separately and then taking 1 ml of the incubated peptone water of each sample, mixing them well and then take 0.1ml of the mixture and inoculate the modified Rappaport-Vassiliadis semisolid medium plates (MSRV). (harmonised sampling method to verify C&D processes).
These samples must be analysed in authorised laboratories in the framework of the national Salmonella monitoring and control programmes.
The detection methods used must be the same as for the other samples under the PNCS. The results for the same must be recorded using the MAPA computer application for own checks.
The samples must be recorded alongside the samples for the outgoing flock. The sampling sheet for own checks must be used when sending such samples to the laboratory. The competent authorities shall check the suitability of the cleaning, disinfection and depopulation measures adopted in the hen houses and, where appropriate, will authorise installations to be occupied by new animals.
If there is a positive result (we detect Salmonella spp.), cleaning and disinfection should be repeated.

f) Adequate measures must be taken to prevent the transmission of Salmonella spp through drinking water.
g) The appropriate measures must be taken to prevent the presence of Salmonella spp in raw materials and feedingstuffs. Specifically, the manufacturer or supplier of feed to the holding must guarantee that testing for Salmonella has been carried out and make express provision for such tests in the relevant HACCP system. The checks must include analysis of the corresponding samples, which will be made available to the health managers of the holdings receiving the feed. The veterinarian responsible for the holding may assist with the interpretation of the results of the analysis.
h) Adequate training courses must be given to workers and appropriate health checks must be carried

out to detect possible contamination of workers on the holding with any of the five Salmonella serotypes if the bacterium is detected in animals.
i) Suitable health checks must be carried out to detect the possible source or sources of Salmonella contamination where the bacterium has been detected in animals or if this emerges from the epidemiological investigation.
j) Appropriate vaccination programmes must be carried out where necessary.
k) Appropriate sampling and analyses are carried out to detect Salmonella spp.
l) Adequate measures must be taken to ensure the traceability of eggs produced in accordance with the legislation in force.
m) Adequate measures must be adopted if positive cases of salmonellosis involving any of the five Salmonella serotypes occur.
n) Appropriate measures must be taken to ensure the proper management of by-products of animal origin not intended for human consumption.

b. Routine official **sampling scheme when FBO sampling takes place at the hatchery**: EU minimum requirements are implemented i.e. :

If the EU target is achieved for more than 2 years, the CA has decided to implement the derogation of point 2.1.2.3 of Annex to Regulation (EC) No 200/2010 and therefore the EU minimum requirement for official sampling are once a year at the hatchery and once a year on the holding during the laying phase.

☐*yes* ☒*no*

If no, the EU minimum requirements for official sampling are implemented as follows:

- every 16 weeks at the hatchery
- twice during the laying phase at the holding (within four weeks at the beginning, within eight weeks before the end), and
- at the holding each time samples taken at the hatchery are positive for target serovars

☐*yes* ☒*no*

If no please explain. Indicate also : 1) if additional official sampling going beyond EU minimum requirements is performed, 2) who is taking the official samples.

Samples are not taken in incubators in Spain.

c. Routine official **sampling scheme when FBO sampling takes place at the holding**: EU minimum requirements are implemented i.e. :

If the EU target is achieved for more than 2 years, the CA has decided to implement the derogation of point 2.1.2.3 of Annex to Regulation (EC) No 200/2010 and therefore the EU minimum requirement for official sampling are twice during the laying phase at the holding.

☒*yes* ☐*no*

If no, the EU minimum requirements for official sampling are implemented as follows:

- Three times during the laying phase at the holding (within four weeks at the beginning, within eight weeks before the end and a third one in between)

☒*yes* ☐*no*

If no, please explain. Indicate also : 1) if additional official sampling going beyond EU minimum requirements is performed, please describe, 2) who is taking the official samples

Official samples will be taken by the qualified or authorised official veterinarian, or in some cases under veterinary supervision by other sufficiently trained authorised personnel. A minimum of three separate official checks on all of the flocks on all holdings with more than 250 birds must be carried out on three occasions during the production cycle:
• The first within four weeks of the transfer to the laying unit;
• The third towards the end of the laying phase, not earlier than eight weeks before the end of the production cycle;
• The second official analysis must be carried out during the productive period at an appropriate interval from the other two.
In addition, sampling by the competent authority shall take place whenever the competent authority considers it appropriate.
Given that the Community target has been reached at national level for at least two consecutive calendar years in Spain, the competent authority may replace the routine samplings by two samplings on the holding, on any two occasions with sufficient time between each other during the production cycle.
It falls to each Autonomous Community to decide whether or not to make use of this exemption. In Spain, most of the Autonomous Communities have decided to make use of it.
During sampling, all the data necessary to identify the sample and the flock from which it comes will be collected and will comprise at least the data set out in the sampling sheet for official checks.
Sampling performed by the competent authority may replace sampling on the initiative of the food business operator (own check).
All data and information gathered on holdings on which official sampling has been performed (SEE THE SAMPLING SHEET FOR OFFICIAL CHECKS and the BIOSECURITY SURVEY) and the laboratory results shall be recorded in a dedicated computer application developed for the national programme for the control

of Salmonella.

Sampling protocol it is the same as the protocol described in point 7 of the programme (sampling in adult breeding flocks) (clarify the protocol applied).

Other official samples
Whenever the competent authority deems it necessary, official samples of animal feed and drinking water and environmental samples may be taken to confirm the effectiveness of cleaning and disinfection measures.
If necessary, it may be requested t he results of laboratory analyses of the worker/s in charge of the animals, or anyone who can be considered as a risk (this text shall be added because it was requested by the sector), in order to determine whether there are any Salmonella spp. carriers among them.

d. If confirmatory samples taken at the holding (after positive results at the hatchery, or suspicion of false positivity on FBO samples taken on the holding) are negative, please describe the measures taken:

- [x] Testing for antimicrobials or bacterial growth inhibitors (at least 5 birds per house) and if those substances are detected the flock is considered infected and eradication measures are implemented (annex II.C of Regulation (EC) No 2160/2003)
- [ ] Other official samples are taken on the breeding flock; if positive, the flock is considered infected and eradication measures are implemented, if negative, all restrictive measures are lifted
- [ ] Other official samples are taken on the progeny; if positive, the flock is considered infected and eradication measures are implemented, if negative, all restrictive measures are lifted
- [ ] None of these measures

Comments - Describe also if any other measures are implemented*(max. 32000 chars) :*

e. **Official confirmatory sampling** (in addition to the confirmatory samples at the holding which are systematically performed if FBO or official samples are positive at the hatchery):

Breeding flocks of Gallus gallus

After positive official samples at the holding	☐ Always ☒ Sometimes (criteria apply) ☐ Never
After positive FBO samples at the holding	☐ Always ☒ Sometimes (criteria apply) ☐ Never

When official confirmatory sampling is performed, additional samples are taken for checking the presence of antimicrobials:

☒ Always ☐ Sometimes ☐ Never

Please insert any comments. Describe the criteria used to determine if confirmatory sampling is performed. Indicate also which samples (if any) are taken to check the presence of antimicrobials.

In exceptional cases, and with a view to ruling out false positives or false negatives for samples taken as part of official controls or own checks, the competent authority may decide to carry out confirmatory analyses:
i) by taking 5 faeces samples or 5 pairs of boot swabs and 2 dust samples of 250 millilitres containing at least 100 grams of dust collected from various locations distributed throughout the shed; dust may also be collected from a surface of at least 900 cm2, or 5 faeces samples or 5 pairs of boot swabs and two additional faeces or boot swab samples may be collected; however, a sub-sample of 25 grams must be collected of each faecal material and dust sample for analysis; all samples must be analysed separately,
or ii) bacteriological investigation of the caeca and oviducts of 300 birds,
or iii) bacteriological investigation of the shell and the content of 4 000 eggs from each flock, in pools of maximum 40 eggs.

In addition to the set arrangements above, the competent authority will check that there has been no use of antimicrobials that might affect the results of the sampling analyses.
Whenever confirmatory testing is conducted, additional samples can be collected for the possible testing of antimicrobials or bacterial growth inhibitors as follows: birds shall be taken at random from within each poultry house of birds on the holding, normally up to five birds per house, unless the competent authority deems it necessary to sample a higher number of birds.
Additionally, samples of feed and water can be taken to determine whether the results of the confirmatory test may have been affected by the use of antimicrobials.

If antimicrobials or bacterial growth inhibitors are detected, the Salmonella infection shall be considered to be confirmed.

Breeding flocks of Gallus gallus

There is a national protocol with the minimum criteria for authorizing a confirmatory sampling requested by the FBO, that include terms of type of production, epidemiological health situation and health history of the farm. Additionnal information can be found in the attached confirmatory protocol.

1	2	3	4
For routine samples taken at the holding	No of flocks positive to SE / ST	Out of the flocks in column 2, No of cases where official confirmatory samples[3] were taken	Out of the cases in column 3, No of cases where confirmatory samples were negative
FBO samples[1]	5	3	2
Official samples[2]	1	1	1

1 Reg 200/2010, point 2.2.2.1 of the Annex

2 Reg 200/2010, point 2.2.2.2 of the Annex

3 Reg 200/2010, point 2.2.2.2.c of the Annex

What happened to the flocks counted under 4 (re checked for the presence of Salmonella (on the progeny? on the same flock)? Checked for the presence of antimicrobials?) (max. 32000 chars) :

In 2021, 4 confirmatory test were made and 3 flocks resulted negative and the infection of the flock with target serovars of Salmonella was discarded.
In 2 of the flocks where confirmatory tests were negative the birds followed the productive cicle with the correlative routine sampling of the FBO and Official samples according to the EU regulation and the national programme, until the end of the productive period with the slaugthering of the birds. The premises were cleaned and disinfected and disinsected and before entering new birds it was made the sampling for verification of cleaning and disinfection, with negative results.
In the other flock with negative confirmatory sampling, the birds were slaughtered and the premises cleaned and disinfected and verified by the proper samples, with negative results.

f. Article 2 of Regulation (EC) No 1177/2006 (**antimicrobials** shall not be used as a specific method to control *Salmonella* in poultry): please describe the official controls implemented (documentary checks, sampletaking) to check the correct implementation of this provision (at the holding and at the hatchery). For samples please describe the samples taken, the analytical method used, the result of the tests.

(max. 32000 chars) :

The checks made by the competent authorities (laboratory tests or documentary checks on the records of the holding) must guarantee that no antimicrobial medicinal products that might affect the result of analyses have been used.
In addition to the sampling provided for, when appropriate a random sample of birds may be taken within each shed housing birds on a holding, usually of up to five birds per flock unless the competent authority considers it necessary to include a greater number of birds in the sampling.

The examination shall consist of a test, using accredited techniques to dectect the effect of bacterial growth inhibitors or antimicrobials.
Samples of feed and water may be taken simultaneously with the aim of detecting and quantifying the quantity of antimicrobials if necessary.
Where the presence of the Salmonella serotypes covered by the programme is not detected but antimicrobials or bacterial growth inhibitory effects are detected it shall be considered and accounted for as an infected flock for the purpose of the Union target.
These samples ,in the framework of the SNCP, shall not take in triplicate notwithstanding that these actions can be combined with other programs in which these samples in triplicate are necessary.
If, from this action, derive measures related to the national plan of investigation of residues of veterinary drugs, it will take the appropriate actions, according to the aforementioned regulations.

16. Salmonella vaccination

- [x] Voluntary
- [ ] Compulsory
- [ ] Forbidden

Use of *Salmonella* vaccines is in compliance with provisions of Article 3 of Regulation (EC) No 1177/2006.

Comments - If performed please describe the vaccination scheme (vaccines used, vaccines providers, target flocks, number of doses administered per bird, etc) (max. 32000 chars) :

Vaccination shall be carried out in accordance with Article 3 of Regulation (EC) No 1177/2006.
Vaccination of breeding hens is not mandatory, but in the event that it is carried out out, only vaccines with prior marketing authorisation from the Spanish Medical and Health Products Agency or the European Commission in accordance with Regulation (EC) No 726/2004 may be used. Where one of the --five-- ++three types of Salmonella (SE, ST, SMT, SI)++(this text shall be removed and added because of updating) is confirmed on heavy breeder holdings, the above-mentioned measures at least shall be adopted and, in addition, the next batch of birds introduced must be pullets vaccinated with authorised vaccines or autovaccines in accordance with the legislation in force, before commencing the laying stage. Once vaccination has been carried out, at least the following information will be entered in the register of treatment with medicinal products: date of vaccination, name of the vaccine(s) administered, type of vaccine(s) administered, quantity (number of doses and quantity of each dose), name and address of the supplier of the medicinal product and identification of the batch of animals treated.
Vaccine use must also be recorded using a computer application.

17. System for **compensation to owners** for the value of their birds slaughtered or culled and the eggs destroyed or heat treated.

Describe the system for compensation to owners. Indicate how improper implementation of biosecurity measures can affect the payment of compensation (max. 32000 chars) :

In cases where birds are subjected to compulsory slaughter, the owners of the birds will be entitled to compensation, provided that they have complied with the animal health legislation in force.
The scales for compensation are fixed by the Ministry of Agriculture, Fisheries and Food following consultation with the Autonomous Communities. The above scales are public and are included in Royal Decree 823/2010 of 25 June 2010, laying down the scales of compensation for the compulsory slaughter of animals covered by the national control programmes for Salmonella in breeding and laying flocks of Gallus gallus and breeding turkey flocks.
The age of the birds for compensation purposes shall be considered to be their age when the competent authority ordered the compulsory slaughter.

18. Please describe the official procedure to test, after the depopulation of an infected flock, the **efficacy of the disinfection** of a poultry house (number of samples, number of tests, samples taken, etc...)

(max. 32000 chars) :

Once the shed housing the infected flock has been depopulated, efficient and thorough cleaning (including complete removal of the bedding and excrement) shall be undertaken, followed by disinfection, insect removal and rat extermination. The above tasks shall be performed using properly authorised and registered products. As soon as sufficient time has elapsed after disinfection, environmental samples shall be taken to check the effectiveness of the cleaning and disinfection process and the absence of Salmonella spp. in the environment.

The competent authorities shall check the suitability of the cleaning, disinfection and depopulation measures adopted in the hen houses and, where appropriate, will authorise installations to be occupied by new animals.

To verify cleaning and disinfection, two or more moistened fabric swabs of at least 900 cm2 per swab, moistened using appropriate diluents (such as 0,8 % sodium chloride, 0,1 % peptone in sterile deionised water, sterile water or any other diluent approved by the competent authority, shall be used to swab as large a surface area in different points in the house (floor, walls, feeding equipments, watering equipments, belts, pilars, water and feeding pipes, scrapers and any other difficult point to clean and disinfect).
Samples can be pooled to perform and single culture, or by enriching the peptone water separately and then taking 1 ml of the incubated peptone water of each sample, mixing them well and then take 0.1ml of the mixture and inoculate the modified Rappaport-Vassiliadis semisolid medium plates (MSRV).
These samples must be analysed in authorised laboratories in the framework of the national Salmonella monitoring and control programmes.
The detection methods used must be the same as for the other samples under the PNCS.
The results for the same must be recorded using the MAPA computer application for own checks.
The samples must be recorded alongside the samples for the outgoing flock.
The sampling sheet for own checks must be used when sending such samples to the laboratory.
If there is a positive result (we detect Salmonella spp.), cleaning and disinfection should be repeated.

The premises shall not be restocked for 12 days after completion of the cleaning, disinfection, rat extermination and if necessary insect removal processes. Repopulation may take place only if the

environmental analyses carried out in accordance with the programme are satisfactory, and if biosecurity measures considered inadequate or deficient by the competent authority have been properly corrected. Notwithstanding the above, in those cases where the results of those tests prove the effectiveness of the cleaning and disinfection undertaken, the waiting period may be reduced to a minimum of 7 days.

B. General information

1. Structure and organisation of the **Competent Authorities** (from the central CA to the local CAs)

Short description and/or reference to a document presenting this description (max. 32000 chars) :

For the purposes of this programme, the competent authorities shall be those of the Autonomous Communities and the General State Administration responsible for animal health matters. The General Subdirectorate for Animal Health and Hygiene of the Ministry of Agriculture, Fisheries and Food (MAPA) is responsible for developing and coordinating this monitoring and control programme and for making any necessary amendments, particularly in the light of the data and results obtained; it shall liaise with the Commission, summarising the data and results obtained for communication to the Commission and reporting on the development of the disease. Royal Decree 1440/2001 of 21 December 2001 setting up the veterinary health warning system created the "National Veterinary Health Warning System Committee" (a diagram of the Health Warning System Network (RASVE) is enclosed), which is responsible for studying and proposing measures to prevent, control, combat and eradicate diseases covered by national programmes. Its tasks were reinforced by Law No 8/2003 on animal health. This committee is attached to the Ministry of the Agriculture, Fish and Food (MAPA), and its members represent all the Autonomous Communities and the Ministry of Health for zoonoses. Its tasks include the following:
a) Coordinating animal health actions across the different administrations.
b) Studying measures for preventing, controlling, combating and eradicating the diseases covered by the national programmes.
c) Monitoring the development of the epidemiological situation with regard to animal diseases at national, European and international level.
d) Proposing relevant measures.
This national committee could agree to set up a consultative committee on avian salmonellosis, which would be attached to it, and would include members of the most representative organisations and associations in this sector in Spain, and may also include the professional association of veterinary officers. The role of this consultative committee would be to advise the Committee when requested to do so and also to put any relevant issues to it for consideration.

2. **Legal basis** for the implementation of the programme

(max. 32000 chars) :

The measures included in this monitoring programme for application when Salmonella is detected

comply with the requirements of parts C and E of Annex II to Regulation (EC) No 2160/2003 of the European Parliament and of the Council, and are implemented in accordance with Commission Regulation (EC) No 200/2010, including requirements for detection tests (type of samples, frequency of sampling, preparation of samples, laboratories, methods of analysis and notification of results).

3. Give a short summary of the outcome of the **monitoring of the target *Salmonella* serovars** (SE, ST) implemented in accordance with Article 4 of Directive 2003/99/EC (evolution of the prevalence values based on the monitoring of animal populations or subpopulations or of the food chain).

(max. 32000 chars) :

Since 1993, Salmonella monitoring and control in Spain has been conducted in accordance with Council Directive 92/117/EEC — repealed by Directive 2009/99/EC — concerning measures for protection against specified zoonoses and specified zoonotic agents in animals and products of animal origin in order to prevent outbreaks of food-borne infections and intoxications. The monitoring and control have focused on S. Enteritidis and S. Typhimurium. Data on breeding flocks of Gallus gallus were monitored and collected throughout 2004 on the basis of instructions given at Community level in order to meet the target for the reduction of prevalence laid down in Regulation (EC) No 2160/2003 of the European Parliament and of the Council on the control of Salmonella and other specified food-borne zoonotic agents. The data obtained from the study showed prevalence of the five serotypes (SE, ST, SH, SV, SI) in the production phase to be 16.6 %, rising to 20.3 %for Salmonella spp. The evolution of prevalence of the monitored Salmonella serotypes in flocks of breeding hens of the species Gallus Gallus was as shown below, with the most prevalent target serotypes being S. monophasic, Typhimurium and SV, followed by SE, ST and SI (a file containing the evolution of prevalence is enclosed):

4. System for the registration of holdings and identification of flocks

(max. 32000 chars) :

Legislative measures and provisions concerning the registration of livestock holdings
The requirement to register livestock holdings in Spain stems primarily from Article 39 of Law 8/2003 of 24 April 2003, the Animal Health Act. More specifically, and where poultry farming is concerned, the requirement to register holdings is regulated by the following instruments: Royal Decree 479/2004, of 26 March 2004, establishing and regulating a general register of livestock holdings. Covers all livestock species. Royal Decree 1084/2005 of 16 September 2005 establishing regulations for poultry farming for meat. Applies to holdings where poultry birds are reared or kept for meat, excluding holdings where birds are kept for own consumption, as defined in Article 2.b. Measures and applicable legislation as regards the identification of animals
The programme shall cover breeding poultry flocks, since individual animals are not identified. For the purposes of the programme an epidemiological unit shall be considered to be a breeding flock, defined as all poultry of the same health status kept on the same premises or within the same enclosure; in the case of housed poultry, this includes all birds sharing the same airspace, in accordance with Article 2(3) (b) of Regulation (EC) No 2160/2003 of the European Parliament and of the Council. Flocks of breeding hens shall be identified individually. To identify the flocks on a holding the REGA code will be used, consisting of a capital letter corresponding to the shed (this letter must be written on the entrance door

to the shed) and the date of entry of the birds into that shed, in the format mmyyyy. REGA+ SHED (CAPITAL LETTER)+ DATE OF ENTRY OF BIRDS (mmyyyy)

5. System to monitor the implementation of the programme.

(max. 32000 chars) :

Taking account of the structure and organisation of the Spanish State, the General State Administration — represented by the Subdirectorate-General for Animal Health and Hygiene and Traceability of the Ministry of Agriculture,Fish and Food (MAPA) is responsible for developing and coordinating this monitoring and control programme and for making any necessary amendments, particularly in the light of the data and results obtained; it shall liaise with the Commission, summarising the data and results obtained for communication to the Commission; lastly, it is responsible for reporting on the development of the disease. The Autonomous Communities are responsible for the direct implementation and monitoring of the activities to be carried out under the programme. In addition, to facilitate monitoring and follow-up of the data obtained, we have two computer applications for recording information from own checks and official controls. Information from own checks is recorded by the authorised laboratories that analyse own-check samples, and information from official controls is recorded by the official veterinary services of the Autonomous Communities. The information is thus subject to double review: the Autonomous Communities review the information from both applications on their territory, and the Subdirectorate-General for Animal Health and Hygiene and Traceability globally reviews all of the results.
Lastly, we have a monitoring plan for own checks and inspection of own-check laboratories: In order to verify that own checks are being performed correctly, the competent authority will implement the following Monitoring Plan for own checks and inspection of own-check laboratories (document enclosed):
The Official Veterinary Services carry out quality controls on own checks on a percentage of holdings, selected each year in accordance with the following ranked risk criteria: • Holdings where results for the serotypes being monitored were negative in own checks and positive in official controls.
• Holdings where results for the serotypes being monitored were negative in own checks but for which there was a Public Health notification of a positive result.
• Holdings where results for the serotypes being monitored were negative in own checks but positive results were obtained for the LOD in effectiveness checks.
• At random on holdings where results for the serotypes being monitored were negative in own checks and no official controls were carried out.
This will involve 5% of the holdings in each Autonomous Community. If there are fewer than 20 holdings in a Community they will be carried out on at least one farm. The control will involve conducting a survey to verify whether the requirements of the programmes are being met. The Autonomous Community may decide to carry out a site inspection of an own-check sampling exercise. In this case, the own-check sampling must take place in the presence of the official veterinarian who, as an observer, will attempt to identify practices that do not correspond to the procedures for sampling set out in detail in the National Programmes and applicable to own checks. Close attention will be paid to critical aspects of those procedures that could presumably affect the results (such as the use of peptone as an enrichment medium for boot swabs, origin, expiry; representativeness of the sample: number of steps taken and surface area covered; where appropriate, dispersion of the collection of aliquots of faeces to generate sufficient representativeness in pools, etc). The manner and location of storage of the sample when delivered to the laboratory must also be checked, as must compliance with the maximum deadlines set for receipt of the samples. It is very important that before any own checks are carried out

on holdings, and whenever routine official controls are carried out, the holding information recorded in the own-check application is consulted. During this inspection the competent authority must also ask any questions considered necessary and request the necessary documentation on the performance of own checks. The official veterinarian must note down the results of the control in an inspection report. The information in that report, and any other information obtained when tracing the sample until it arrives in the laboratory, will be used by the competent authority to draw up an appraisal report. In the event that any shortcomings are detected, these must be reported to the producer as soon as possible to be corrected immediately for future own checks, without prejudice to any administrative consequences this may have. The competent authority must issue a copy of the report to the party responsible for taking the own-check samples. If the competent authority considers it appropriate, duplicate samples will be taken. One of the samples will be taken by the official veterinarian using his or her own materials. This sample will be retained by the veterinarian and will be sent to an official laboratory together with the sampling sheet. The other sample will be taken by the party responsible for taking the own-check samples, using material provided by that party. It will remain in that party's possession and must be analysed in the same way as any other own-check sample. In those cases where there are significant discrepancies between the results of the official controls and the own checks in the same flock, the competent authority may, if it considers it appropriate, request the strains isolated from the flock in question from the own-check laboratory where they were tested and test them in an official laboratory of the Autonomous Community concerned. Inspections in laboratories will take place in accordance with the document enclosed above. Each Autonomous Community must have inspected all of the laboratories in its territory within two years.

C. Targets

1 Targets related to flocks official monitoring

1.1 Targets on laboratory tests on official samples for year : 2023

Type of the test (description)	Target population	Number of planned tests
Bacteriological detection test	Breeding flocks of Gallus gallus	5 000
Serotyping	Breeding flocks of Gallus gallus	100
Antimicrobial detection test	Breeding flocks of Gallus gallus	50
Test for verification of the efficacy of disinfection	Breeding flocks of Gallus gallus	10

1.2 Targets on official sampling of flocks for year : 2023

Type of the test (description)	Rearing flocks	Adult flocks
Total No of flocks (a)	1 020	1 720
No of flocks in the programme	1 020	1 720
No of flocks planned to be checked (b)	5	1 700
No of flock visits to take official samples (c)	5	2 500
No of official samples taken	35	5 010
Target serovars (d)	SE+ ST + SH +SI + SV	SE+ ST + SH +SI + SV
Possible No of flocks infected by target serovars	3	8
Possible No of flocks to be depopulated	3	8
Total No of birds to be slaughtered/culled	20 000	80 000
Total No of eggs to be destroyed	Text	423 000
Total No of eggs to be heat treated	Text	770 000

Breeding flocks of Gallus gallus

(a) Including eligible and non eligible flocks
(b) A checked flock is a flock where at least one official sampling visit will take place. A flock shall be counted only once even if it was visited serveral times.
(c) Each visit for the purpose of taking official samples shall be counted. Several visits on the same flock for taking official samples shall be counted separately.
(d) Salmonella Enteritidis and Salmonella Typhimurium = SE + ST
Salmonella Enteritidis, Typhimurium, Hadar, Infantis, Virchow = SE+ ST + SH +SI + SV

2.1 *Targets on vaccination for year :* **2023**

Type of the test (description)	Target on vaccination
Number of flocks in the Salmonella programme	1 720
Number of flocks expected to be vaccinated	1 600
Number of birds expected to be vaccinated	14 500 000
Number of doses expected to be administered	50 000 000

E. Financial information

1. Identification of the implementing entities - financial circuits/flows

Identify and describe the entities which will be in charge of implementing the eligible measures planned in this programme which costs will constitute the reimbursment/payment claim to the EU. Describe the financial flows/circuits followed.
Each of the following paragraphs (from a to e) shall be filled out if EU cofinancing is requested for the related measure.

a) Implementing entities - **sampling**: who perform the official sampling? Who pays?
(e.g. authorised private vets perform the sampling and are paid by the regional veterinary services (state budget); sampling equipment is provided by the private laboratory testing the samples which includes the price in the invoice which is paid by the local state veterinary services (state budget))

The official samples are taken by official veterinarians. The cost of sampling is covered by the administrative authorities, in this case the Autonomous Communities.

b) Implementing entities - **testing**: who performs the testing of the official samples? Who pays?
(e.g. regional public laboratories perform the testing of official samples and costs related to this testing are entirely paid by the state budget)

The official samples are analysed in the official laboratories of the Autonomous Communities. The cost of the analyses is covered by the Autonomous Community. The national reference laboratory (NRL, Algete) also carries out serotyping analysis of official samples. To a lesser extent, it also performs isolation and identification analyses. These analyses are paid for by the NRL.

c) Implementing entities - **compensation**: who performs the compensation? Who pays?
(e.g. compensation is paid by the central level of the state veterinary services,
or compensation is paid by an insurance fund fed by compulsory farmers contribution)

The official veterinary services of the Autonomous Communities (ACs) organise compulsory slaughter and are responsible for providing slaughter compensation. The ACs are responsible for financing this. For broiler chickens, slaughter in the case of positive flocks is not compulsory and therefore is not compensated.

d) Implementing entities - **vaccination**: who provides the vaccine and who performs the vaccination? Who pays the vaccine? Who pays the vaccinator?
(e.g. farmers buy their vaccine to the private vets, send the paid invoices to the local state veterinary services which reimburse the farmers of the full amount and the vaccinator is paid by the regional state veterinary services)

The vaccination of breeding hens is voluntary. The private veterinarians working for a Livestock Health Association provide and perform the vaccination for the birds of the holding of the farmer that contract the services of that association.
The administrative authorities may finance the vaccination based on regional grants for the Livestock Health Associations. Regional veterinary services will reimburse these associations after checking the corresponding documents (invoices of purchase, nº of animals vaccinated, nº of doses used, date of vaccination, etc).

e) Implementing entities - **other essential measures**: who implement this measure? Who provide the equipment/ service? Who pays?

Installations are always cleaned and disinfected after the sheds have been emptied. Before repopulating the sheds, cleaning and disinfection must be checked, taking environmental samples. These activities are the responsibility of the food business operators, who pay for them. On some occasions, the competent authority of the ACs also takes samples to check the effectiveness of cleaning and disinfection, in which case the administrative authorities cover the cost.

2. Source of funding of eligible measures

All eligible measures for which cofinancing is requested and reimbursment will be claimed are financed by public funds.

☒*yes*

☐*no*

3. Additional measures in exceptional and justified cases

In the "*Guidelines for the Union co-funded veterinary programmes*", it is indicated that in exceptional and duly justified cases, additional necessary measures can be proposed by the Member States in their application.

If you introduced these type of measures in this programme, for each of them, please provide detailed technical justification and also justification of their cost:

Attachments

IMPORTANT :

1) The more files you attach, the longer it takes to upload them .
2) This attachment files should have one of the format listed here : jpg, jpeg, tiff, tif, xls, xlsx, doc, docx, ppt, pptx, bmp, pna, pdf.
3) The total file size of the attached files should not exceed 2 500Kb (+- 2.5 Mb). You will receive a message while attaching when you try to load too much.
*4) IT CAN TAKE **SEVERAL MINUTES TO UPLOAD** ALL THE ATTACHED FILES. Don't interrupt the uploading by closing the pdf and wait until you have received a Submission Number!*
5) Only use letters from a-z and numbers from 1-10 in the attachment names, otherwise the submission of the data will not work.

List of all attachments

		Attachment name	File will be saved as (only a-z and 0-9 and -_) :	File size
			Total size of attachments :	